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Reidispongiolide A and B, Two New Potent Cytotoxic Macrolides from the New Caledonian Sponge *Reidispongia coerulea*

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Abstract: Two new 26-membered macrolides, reidispongiolide A (1) and B (2), have been isolated from the New Caledonian marine sponge *Reidispongia coerulea* n.gen. n.sp. Lévi and Lévi and their structures elucidated. They are related to sphinxolides previously isolated from an unknown nudibranch and later from the sponge *Neosiphonia superstes*. 1 and 2 co-occurr with sphinxolide B (4) and D (3). These macrolides exhibited potent cytotoxicity against various human carcinoma cells.

The isolation of cytotoxic marine sponge macrolides has been rapidly increasing. They have been isolated from sponges of the genus *Theonella*¹, *Halichondria*², *Mycale*³, *Japsis*⁴ and very recently from a *Spongia* sp.⁵, a *Cinachyra* sp⁶. and *Hyrtios altum*⁷. Previously we described discovery of potent cytotoxic 26-membered macrolides, sphinxolide⁸ and its congeners sphinxolides B-D, in the New Caledonian sponge *Neosiphonia* superstes (fam. Phymathellidae)⁹.

Continued investigation of New Caledonian marine invertebrates for bioactive metabolites resulted in isolation and structural elucidation of two new cytotoxic macrolides, herein named reidispongiolide A (1) and B (2), from the marine sponge *Reidispongia coerulea* (Demospongiae, Lithistida, Phymatellidae) n.gen. and n.sp.



Lévi and Lévi¹⁰, collected in the South of New Caledonia in the region Banc Eponge sea-mount at a depth of 500 m. These compounds are related to sphinxolides⁹, and co-occurr with sphinxolide B (4) and D (3). Like the previous sphinxolides, reidispongiolide A (1) and B (2) are extremely cytotoxic against various human carcinoma cells *in vitro* (Table 1).

RESULTS AND DISCUSSION

The lyophilized sponge (1 Kg) was extracted in Soxhlet with *n*-hexane and CH_2Cl_2 , and then at room temperature with CH_2Cl_2 :MeOH (8:2) and finally with MeOH. The cytotoxic (*Artemia salina* assay) dicloromethane extract (2.4g) was chromatographed on silica gel (flash column, MeOH/CHCl₃ 0-20%) followed by reverse phase $C_{18}\mu$ -bondapak HPLC with 77% MeOH aq. to give reidispongiolide A (1, 83.1 mg) and sphinxolide D (3, 23.9 mg) and with 75% MeOH aq. to give reidispongiolide B (2, 2.6 mg) and sphinxolide B (4, 4.8 mg).

By means of positive fast atom bombardment mass spectrometry (FABMS), which showed the largest ion peaks at m/z 980 (M+Na)⁺ and 958 (M+H)⁺ and by ¹³C NMR data (Table 2), the molecular formula of the major macrolide reidispongiolide A (1), $[\alpha]_D$ =-4.8°, was shown to be $C_{s4}H_{s7}O_{13}N$, which is 16 mass units less than sphinxolide D (3). ¹H and ¹³C NMR spectra of 1 closely resembled those of sphinxolides and particulary that of sphinxolide D (3). The ¹H NMR (Table 2) spectrum of 1 showed seven methoxyl singlets, six methyl doublets,

Table 1. In vitro cytotoxic activity (IC_{so} in µg/ml) of reidispongiolide A (1) and B (2).

Tumor cells	1	2	6-mercaptopurine
NSCLC-N6	0.07	0.05	0.76
P388/Dox	0.01	0.02	0.25
P388	0.16	0.06	0.70
КВ	0.10	0.06	0.54
HT29	0.04	0.04	0.87

a) NSCLC-N6: human bronchopulmunary non-small-cell-lung-carcinoma. P388: murine leukemia. P388/Dox : murine leukemia expressing the multi-drug resistance gene mdr, expecially towards doxorubicine. KB : human nasopharyngeal carcinoma. HT29 : human colon carcinoma. Results corresponding to the averages of three to five experiments.

one vinylic methyl singlet and, like sphinxolides, restricted rotation about the N-methyl formyl terminus gave rise to doubled signals for H-37, N-Me, H-36, H-35 and 33-OMe. Interpretation of 2D-COSY (Fig. 1) disclosed that C_2-C_8 , $C_{25}-C_{30}$ and $C_{32}-C_{36}$ portions were those of the 8-demethoxysphinxolides (3 and 4) and revealed the ¹H coupling network $C_{10}-C_{25}$ in which C-10 was a methylene (δ 2.29 and 2.19 ppm). Thus, reidispongiolide A (1) was presumed to be the 10-deoxysphinxolide D. The C-10 signal at 63.0 ppm in 3 was replaced in 1 by a methylene carbon (DEPT) at 31.2 ppm, whereas the C-11 and C-12 signals in 1 were found shifted highfield to 79.3 ppm (82.3 ppm in 3) and downfield to 39.2 ppm (35.7 ppm in 3), respectively, as expected for the removal of the hydroxyl at C-10. The remaining signals were almost identical in both spectra apart small variations for the signals of carbons around C-10, *viz.* at C-7 (76.9 in 1 *vs.* 78.0 ppm in 3), C-8 (40.8 in 1 *vs.* 39.2 ppm in 3), C-9 (157.6 in 1



Fig.1 Expanded region of 2D-COSY (500 MHz, CDCl₃) of reidispongiolide A (1)

	1		2		
Position	ð _u	ð,	ðu	ð,	
1	- "	166.7	-	166.7	
2	5.79 d (15.3)	120.5	5.82 d (15.3)	120.6	
3	7.50 dd (15.3,11.8)	140.2	7.50 (15.3,11.8)	140.2	
4	6.00 d (11.8)	126.1	6.00 d (11.8)	125.7	
5	-	145.5	-	145.5	
6	2.08,2.48 dd	44.5	2.06,2.46 dd	44.4	
7	3.45 m	76.9	3.46 m	78.2	
8	2.20 dd (3.16,14.7),2.30 m	40.8	2.23 dd (3.16,14.7),2.37 m	40.8	
9	-	157.6	•	157.6	
10	2.19,2.29	31.2	2.20,2.35	30.6	
11	4.22 m	79.3	4.22 m	78.8	
12	1.78 dd	39.2	1.92 dd	39.2	
13	3.20 m	77.8	3.26 m	78.3	
14	1.72.1.89 m	33.4	1.63.1.89 m	34.5	
15	3.48 m	79.2	3.50 m	78.8	
16	5.18 dd (15.4.8.5)	129.9	5 26 dd (15 4 8 5)	131.0	
17	5.52 dd (15.4.7.8)	139.2	546 dd (15478)	139.0	
18	2.32 dd	37 5	2 28 dd	41 2	
19	3.08 m	80.6	3.40 m	73.2	
20	1 23 1 66 m	36.6	1 45 1 58 m	397	
20	3.38 m	JU.U 79.9	2.60 m	30.7 70.9	
21	5.00 M	120.2	5.09 m 5.10 dd	131 4	
22	5 42 dd (15 1 30 0)	138.6	5.17 dd	131.4	
20	2 42 dd (13.1,10.0)	136.0	3.51 uu 2.49 d.4	137.4	
24	5 12 44	40.0	2.47 uu 5 10 44	40.0	
25	1 90 44	13.3	5.10 dd	13.5	
20		30.4 97.1	1.95 dd	30.5	
27	2.66 dd	8/.1	2.70 dd	87.2	
20	1.05 III 1.22 1.65 m	34.3	1.08 m	34.4	
29	1.32,1.65 m	23.2	1.43,1.71 m	23.3	
30	2.45,2.55 m	40.8	2.45,2.52 m	40.8	
31	-	213.5		213.5	
32	2.70 d	49.0 (48.8)*	2.72 d	48.9 (49.0)°	
33	3.41 m	82.2	3.47 m	82.7	
34	2.11,2.42 m	30.6 (30.3)*	2.16,2.42 m	30.4 (29.8) ^a	
35	5.11-5.06° m	105.4 (107.1)*	5.11-5.06 ^a m	105.4 (107.1) ^a	
36	6.47-7.18° d (15.0)	130.5 (126.3) ^a	6.5-7.18° d (15.0)	130.3 (126.3) ^a	
37	8.28-8.02 ^a s	162.1 (160.8) ^a	8.28-8.02" s	162.0 (160.0)ª	
38	1.87 s	17.9	1.90 s	17.8	
39	5.83 s	117.4	5.90 s	118.3	
40	-	164.0	-	164.1	
41	1.10 d (6.9)	9.8	1.10 d (6.9)	11.0	
42	0.84 d (6.9)	14.0	0.85 d (6.9)	14.2	
43	1.03 d (6.9)	17.4	1.03 d (6.9)	17.8	
44	0.91 d (6.9)	9.9	0.91 d (6.9)	9.9	
45	0.97 d (6.9)	17.5	0.98 d (6.9)	17.5	
46	0.97 d (6.9)	12.7 (12.8) ^a	0.98 d (6.9)	12.7 (12.8)*	
47	3.03-3.08 ^a s	27.5 (32.9)°	3.03-3.08° s	27.5 (33.0) ^a	
OMe-7	3.26 s	56.8	3.24 s	56.0	
OMe-13	3.13* s	55.5*	3.23 s	55.6	
OMe-15	3.20* s	55.6*	3.30 s	56.0	
OMe-19	3.15 s	57.4	-	-	
OMe-21	3.27 s	57.0	3.31 s	56.9	
OMe-27	3.30 s	61.5	3.34 s	61.6	
OMe-33	3.30-3.25° s	57.6	3.36-3.29 [*] s	56.8	

Table 2. ¹H and ¹³C NMR data for reidispongiolide A (1)^{he} and B (2)^e (CDCl₂)

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*Signals for minor conformer. The assignents were aided by COSY, HETCOR^b and DEPT^e experiments. The coupling constants are given in Hz and enclosed in parentheses.

vs. 156.4 ppm in 3) and C-39 (117.4 in 1 vs. 119.9 ppm in 3). Thus the new reidispongiolide A can be defined as the 10-deoxysphinxolide D (1).

Reidispongiolide B (2), $[\alpha]_{D}$ =+3.5°, was smaller than reidispongiolide A (1) by a CH₂ unit, FABMS, *m/z* 966 (M+Na)⁺. Since the 'H NMR spectrum showed signals for only six methoxyl groups, reidispongiolide B was presumed to be a des-O-methyl reidispongiolide A.Analysis of the COSY spectrum revealed that in C₁₀-C₂₅ segment C-10 was a methylene (δ 2.35 and 2.20 ppm) as in 1, and H-19 was shifted downfield by 0.32 ppm (δ 3.40 in 2 vs. 3.08 ppm in 1), thus indicating that 19-methoxyl group was replaced by a free hydroxyl group on this carbon. In confirmation, C-19 signal at 80.6 ppm in 1 was replaced by a signal upfield shifted to 73.2 ppm in 2. A comparison of the spectral data with those of sphinxolide B (4), definitively estabilished the structure of reidispongiolide B as 10-deoxysphinxolide B (2).

Like the other previous sphinxolides, reidispongiolides, which exist as a mixture of two slowly interconverting isomers, are resistant at attempt of crystallization and their stereochemistry remain to be determined.

EXPERIMENTAL

General Methods.

For general methods see : M. Valeria D'Auria et al., *Tetrahedron*, **1993**, 49, 8657-8664. Animal Collection and Preliminary Experiments.

Reidispongia coerulea Lévi and Lévi (Demospongiae, Lithistida, Phymatellidae) was collected during the dredging campaigns (1987, 1989) of the ORSTOM-CNRS, Programme "Substances Marines d'Intérest Biologique (SMIB)" in the South of New Caledonia (Banc Eponge sea-mount, on the Norfolk Ridge) at depth of 500-515 m. Taxonomic identification was performed by Lévi and Lévi¹⁰ at the ORSTOM Centre de Nouméa where reference specimens are on file (reference 1407). Preliminary assays for cytotoxic (KB cells and P388 leukemia cells) and antifungal activities (*Fusarium oxysporum, Phythophthora hevea, Penicillium digitatum, Botrytis cinerea, Pyricularia oryzae* and *Helminthosporium sativa*) showed a marked activity of chloroformic extract. *Extraction.*

The organisms were freeze dried and the lyophilized material (1 Kg) was extracted in Soxhlet with n -hexane and CH_2Cl_2 , then with CH_2Cl_2 :MeOH 8:2 (3x1L) and with MeOH (3L) at room temperature. The dichloromethane extract was filtered and concentrated under reduced pressure to give 2.4 g of a yellow cytotoxic oil (*Artemia salina* assay).

Isolation.

The crude dichloromethane extract was chromatographed by MPLC on a SiO₂ column (50 g) using a solvent gradient system from CHCl₃ to CHCl₃:MeOH 8:2. Fractions eluted with CHCl₃:MeOH 99:1 (340mg) were further purified by HPLC on a Waters C-18 μ -Bondapak column (7.8 mm i.d. x 30 cm) with 77% MeOH aq. to give 83.1 mg of reidispongiolide A (1, t_r=12.8 min.) and 23.9 mg of sphinxolide D (3, t_r=18.0 min) and with 75% MeOH aq. to give 2.6 mg of reidispongiolide B (2, t_r=19.6 min.) and 4.8 mg of sphinxolide B (4, t_r=13.2).

Compound 1 m/z 980 (M+Na)⁺; $[\alpha]_{D}$ =-4.8°

Compound 2 m/z 966 (M+Na)⁺; $[\alpha]_{D} = +3.5^{\circ}$

Compound 3 m/z 996 (M+Na)⁺; $[\alpha]_{D} = -3.2^{\circ}$

Compound 4 m/z 982 (M+Na)⁺; $[\alpha]_{D}$ =+2.8°

Determination of biological activity.

For cytotoxic assays see : M.V. D'Auria et al. Tetrahedron 1993, 49, 8657-8664.

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REFERENCES

- (a) Carmely, S.; Kashman, Y. Tetrahedron Lett. 1985, 26, 511-514. (b) Kobayashi, M.; Tanaka, J.; Katori, T.; Matsuura, M.; Kitagawa, I. Tetrahedron Lett. 1989, 30, 2963-2966. (c) Kobayashi, M.; Tanaka, J.; Katori, T.; Matsuura, M.; Yamashita, M.; Kitagawa, I.Chem. Pharm. Bull. 1990, 38, 2409-2418. (d) Kobayashi, M.; Tanaka, J.; Katori, T.; Kitagawa, I. Chem. Pharm. Bull. 1990, 38, 2960-2966. (e) Doi, M.; Ishida, T.; Kobayashi, M.; Kitagawa, I. J. Org. Chem. 1991, 56, 3629-3632.(f) Tanaka, J.; Higa, T.; Kobayashi, M.; Kitagawa, I. Chem. Pharm. Bull. 1990, 38, 2967-2970.
- (a) Kernan, M.R.; Faulkner, D.J. Tetrahedron Lett. 1987, 28, 2809-2812. (b) Kernan, M.R.; Molinski, T.F.; Faulkner, D.J. J. Org. Chem. 1988, 53, 5014-5020.
- (a)Fusetani, N.; Yasumuro, K.; Matsunaga, S.; Hashimoto, K. Tetrahedron Lett. 1989, 30, 2809-2812.
 (b)Northcote, P.T.; Blunt, J.W.; Munro, M.H.G. Tetrahedron Lett. 1991, 32, 6411-6414.
- 4. Kobayashi, J.; Murata, O.; Shigemori, H.; Sasaki, T. J. Nat. Prod. 1993, 56, 787-791.
- Pettit, G.R.; Cichacz, Z.A.; Gao, F.; Herald, C.L.; Boyd, M.R.; Schmidt, J.M.; Hopper, J.N.A.; J. Org. Chem. 1993, 58, 1302-1304.
- 6. Fusetani, N.; Shinoda, K.; Matsunaga, S.; J. Am. Chem. Soc. 1993, 115, 3977-3981.
- 7. Kobayashi, M.; Aoki, S.; Sakai, H.; Kawazoe, K.; Kihara, N.; Sasaki, T.; Kitagawa, I. Tetrahedron Lett. 1993, 34, 2795-2798.
- 8. Guella, G.; Mancini, I.; Chiasera, G.; Pietra, F. Helv. Chim. Acta 1989, 72, 237-246.
- D'Auria, M.V.; Gomez Paloma, L.; Minale, L.; Zampella, A.; Verbist, J.F.; Roussakis, C.; Debitus, C. Tetrahedron, 1993, 49, 8657-8664.
- 10. Lévi, C.; Lévi, P.; Bull. Mus. natn. Hist. nat. Paris, 4e sér, A, 1988, 10, 241-263.

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